Under centrifugation, i.e., the failure to apply forces necessary to cause the cells to form a “button” and a clear supernate, may result in weak or negative reactions. No one speed and time of centrifugation can be recommended which will cover the wide variety of centrifuges available. Each laboratory must calibrate its own equipment and determine the time required at a given speed to achieve the desired result.

BIBLIOGRAPHY


LIMITED EXPRESSED WARRANTY DISCLAIMER

The manufacturer limits the warranty to the test kit, in as much as that the test kit will function as an in vitro diagnostic assay within the limitations and specifications as described in the product instruction-manual, when used strictly in accordance with the instructions contained therein. The manufacturer disclaims any warranty expressed or implied including such expressed or implied warranty with respect to merchantability, fitness for use or implied utility for any purpose. The manufacturer’s liability is limited to either replacement of the product or refund of the purchase price of the product and in no case liable to for claim of any kind for an amount greater than the purchase price of the goods in respect of which damages are likely to be claimed. The manufacturer shall not be liable to the purchaser or third parties for any injury, death or economic loss, howsoever caused by the product in the use or in the application there of.

1. Label ten small test tubes.

2. Prepare a 5% suspension in Bovine Albumin of appropriate red cells of selected genotype.

3. Prepare a 5% suspension in Bovine Albumin of the patient’s red cells.

4. Make progressively two fold serial dilutions of the patient serum as follows:
   (a) To each tube, except the first, add 0.2ml of serum.
   (b) With a clean autopipette, deliver 0.1 ml. of patient serum to each of the first and second tube.

5. To Tubes 1 through 9, add 0.1 ml. of the albumin suspended selected red cells.

6. To tube 10, add 0.1 ml. of the albumin suspended patient cells.

7. Mix the contents of each tube and incubate at 37°C for a minimum of 15 minutes. If desired, incubation may be extended up to 60 minutes.

8. Centrifuge all tubes and read for agglutination.

9. Perform the antiglobulin test on all tubes that are not already strongly agglutinated.

RESULTS INTERPRETATION

The reciprocal of the dilution in the last tube containing visible agglutination is the end point of titration.

LIMITATION OF PROCEDURE

1. Bovine Albumin will not bring about agglutination of red cells by all IgG blood group antibodies.

2. Red cells that have a positive direct antiglobulin test must not be used for the direct antiglobulin test.

3. Contaminated blood specimen and/or supplementary materials used in the procedures described in this manual may interfere with the test results.

SPECIFIC PERFORMANCE CHARACTERISTICS

Bovine Albumin is known to enhance the reactivity of Rh and some other antibodies. The centrifugal force applied to cell/serum mixture should be the minimum required to produce a “button” of red cells and a clear supernatant. Overcentrifugation, i.e., the application of force in excess, causes the cells to adhere to the bottom of the test tube so that vigorous agitation is necessary before they can be resuspended. During such agitation, weak agglutination, may be dispersed causing a positive reaction to be missed.
**SPECIMEN COLLECTION & PREPARATION**

For indirect antiglobulin test, serum not more than 48 hours old from clotted blood should be used. If plasma is used in the indirect antiglobulin test, complement dependent antibodies may not be detected because Calcium is not available. If delay in testing, store sample at 2-8°C.

**WARNING FOR USERS**

**CAUTION:** ALL THE SAMPLES TO BE TESTED SHOULD BE HANDLED AS THOUGH CAPABLE OF TRANSMITTING INFECTION. NO TEST METHOD CAN OFFER COMPLETE ASSURANCE THAT HUMAN BLOOD PRODUCTS WILL NOT TRANSMIT INFECTION.

1. The use of disposable gloves and proper biohazardous clothing is STRONGLY RECOMMENDED while running the test.
2. In case there is a cut or wound in hand, DO NOT PERFORM THE TEST.
3. Do not smoke, drink or eat in areas where specimens or kit reagents are being handled.
4. Tests are for in vitro diagnostic use only and should be run by competent person only.
5. Do not pipette by mouth.
6. All materials used in the assay and samples should be decontaminated in 5% sodium hypochlorite solution for 30-60 min. before disposal or by autoclaving at 121°C at 15psi for 60 min. Do not autoclave materials or solution containing sodium hypochlorite. They should be disposed of in accordance with established safety procedures.
7. Wash hands thoroughly with soap or any suitable detergent, after the use of the kit. Consult a physician immediately in case of accident or contact with eyes, in the event that contaminated material are ingested or come in contact with skin puncture or wounds.
8. Spills should be decontaminated promptly with Sodium Hypochlorite or any other suitable disinfectant.
9. The reagent contain Sodium Azide as a preservative. If these material are to be disposed off through a sink or other common plumbing systems, flush with generous amounts of water to prevent accumulation of potentially explosive compounds. In addition, consult the manual guideline “Safety Management No. CDC-22", Decontamination of Laboratory Sink Drains to remove Azide salts" (Center for Disease Control, Atlanta, Georgia, April 30, 1976.)

**PROCEDURES AND USES OF BOVINE ALBUMIN**

**A. Compatibility Testing**

See procedure below.

**B. Antibody Detection.**

See procedure below.

**C. Antibody Identification**

For B & C. See direction circular accompanying Anti-Human Serum.

**D. Antibody Titration**

See procedure below.

**E. Control of Rh typing**

For procedure, refer instruction manual of Anti-D (IgM) Monoclonal Antibodies for Slide Test And Modified Test Tube.

**F. D-Testing**

For procedure refer, instruction manual of Anti-D (IgG) Monoclonal and Anti-D (IgM + IgG) Monoclonal Antibodies for Slide test and Modified Tube test.

**REQUIRED SUPPLEMENTARY MATERIALS**

**For tube test method**

1. Test Tubes, 10x75 mm or 12x75mm.
2. Pasteur pipettes.
3. Red Blood Cells (Human) of specific genotypes for anti body detection and for identification Anti-D (IgM) Monoclonal antibodies for slide test and Modified Tube test.
4. Centrifuge
5. Incubator, 37°C
6. Isotonic saline.
8. Coombs Control Cells.

**Additional Material for Titration**

10. 0.2ml. serological pipettes.
11. Red Blood cells (Human) which possesses the antigen specific for the antibody to be titrated.

**SLIDE TEST METHOD**

1. Glass Slides
2. Pasteur pipettes
3. Applicator Sticks
4. Anti-D (IgM) Monoclonal antibodies for Slide Test and Modified Tube Test

**DIRECTIONS FOR USE OF WIDE SPECTRUM COMPATIBILITY TEST**

**MAJOR CROSSMATCH PROCEDURE**

**INITIAL PHASE**

1. For each donor to be cross matched, label two small test tubes. Suggested labelling: S, (for saline) and A (for Albumin)
2. Prepare a 5% suspension of the donor red blood cells in isotonic saline or their own serum or plasma.
3. With a clean Pasteur pipette, add two drops of the recipient serum to each test tube.

4. With a clean pasteur pipette, add one drop of the donor red cells to each test tube.
5. To the Albumin tube only add two drops of Bovine Albumin.
6. Mix well.
7. Centrifuge both tubes. Suggested centrifugation: 15 seconds at 3400 rpm (900-1000rcf) or 1 minute at 1000 rpm (100-125rcf)
8. Examine both tubes macroscopically for hemolysis and/or agglutination.

**INCUBATION PHASE**

9. Incubate the saline tube at room temperature and the Albumin tube at 37°C for a minimum of 15 minutes. If desired, incubation may be extended upto 60 minutes.
10. Centrifuge both tubes.
11. Examine both tubes macroscopically for hemolysis and/or agglutination.
12. Proceed to Antiboglobin Phase with the Albumin tube.

**ANTIGLOBULIN PHASE**

13. Utilizing the albumin tube only, wash the cell/serum mixture thoroughly a minimum of three times with tubes full of Saline. Decant and drain completely after use.
14. Add two drops of Anti Human Serum to the sedimented cells.
15. Mix well and centrifuge
16. Resuspend the cells by gently agitation and examine macroscopically for agglutination. If negative examine with an optical aid.
17. To all negative antiglobulin tests add one drop of Coombs Control Cells. Refer instruction manual of Anti Human Serum for details.

**COOMBS’ CONTROL CELLS**

Dilute Anti-D (IgG) Monoclonal Antibodies for Slide Test and Modified Tube Test 1:10 in isotonic saline. Add an equal volume of this dilution to a 10% cell suspension of Group O Rh1 (D) positive (O+) red blood cells and incubate at 37°C for 30 minutes for sensitization. After incubation, wash the cells thrice with fresh isotonic saline (fill the tube about 50 volumes) and again make 3-4% suspension of sensitized cells in isotonic saline. Add one drop of these Coombs Control Cells suspension to negative antiglobulin tests, mix gently and centrifuge for 15 seconds at 3400 rpm (900-1000rcf) or 1 minute at 1000 rpm (100-125rcf). Agglutination validates the antiglobulin test.

**MINOR CROSS MATCH PROCEDURE**

This procedure is the same as that used for the major side of the Broad Spectrum Compatibility test except that the serum or plasma of the donor is tested against the red cells of the recipient.

**RESULTS INTERPRETATION**

If no hemolysis or agglutination is present through step 16 and the Coombs control cells are agglutinated, the patient and donor may be considered...